

# Multiple Genome Sequences of the Important Beer-Spoiling Species *Lactobacillus backii*

Andreas J. Geissler,<sup>a</sup> Jürgen Behr,<sup>b</sup> Rudi F. Vogel<sup>a</sup>

Technische Universität München, Lehrstuhl für Technische Mikrobiologie, Freising, Germany<sup>a</sup>; Technische Universität München, Bavarian Center for Biomolecular Mass Spectrometry, Freising, Germany<sup>b</sup>

***Lactobacillus backii* is an important beer-spoiling species. Five strains isolated from four different breweries were sequenced using single-molecule real-time sequencing. Five complete genomes were generated, which will help to understand niche adaptation to beer and provide the basis for consecutive analyses.**

Received 17 June 2016 Accepted 6 July 2016 Published 25 August 2016

**Citation** Geissler AJ, Behr J, Vogel RF. 2016. Multiple genome sequences of the important beer-spoiling species *Lactobacillus backii*. *Genome Announc* 4(4):e00826-16. doi:10.1128/genomeA.00826-16.

**Copyright** © 2016 Geissler et al. This is an open-access article distributed under the terms of the [Creative Commons Attribution 4.0 International license](https://creativecommons.org/licenses/by/4.0/).

Address correspondence to Jürgen Behr, [juergen.behr@wzw.tum.de](mailto:juergen.behr@wzw.tum.de).

Beer is a selective environment for the growth of bacteria. Restrictive parameters in beer include ethanol, carbon dioxide, antibacterial hops, and anaerobicity. In addition, beer is characterized by a low pH (3.8 to 4.7) and a selective nutrient content (1–3). Nevertheless, lactic acid bacteria (LAB) of the genus *Lactobacillus* are capable of growing in and spoiling beer. Between 2010 and 2013, *Lactobacillus backii* caused 4.8 to 10% of all beer spoilage incidents in Germany, while spoiled beers are characterized by visible turbidity and slight acidification (4, 5). In order to gain insights into the genomic adaptation of *L. backii* to beer, we sequenced the complete genomes of five brewery isolates with the ability to spoil beer.

Beer spoilage ability was tested as described previously (6). High-molecular-weight DNA was purified from de Man, Rogosa, and Sharpe (MRS) liquid cultures using the Genomic-tip 100/G kit (Qiagen), as described previously (6). Single-molecule real-time sequencing (7) (PacBio RS II) was carried out at GATC Biotech (Konstanz, Germany). An insert size of 8 to 12 kb was selected for library creation, resulting in at least 200 Mb raw data from 1 to 2 SMRT cells (1 × 120-min movies) applying P4-C2 chemistry. Assembly was done with SMRT Analysis version 2.2.0.p2, using the Hierarchical Genome Assembly Process (HGAP) (8), and completed by manual curation (<https://github.com/PacificBiosciences/Bioinformatics-Training/wiki/Finishing-Bacterial-Genomes>). Genomes were

annotated using the NCBI Prokaryotic Genome Annotation Pipeline (PGAP) and the Rapid Annotations using Subsystems Technology (RAST) server (9–11). Pan- and core genomes were calculated using CMG-Biotools and BADGE (6, 12).

Strain characteristics, sequencing statistics, genome information, and accession numbers are listed in Table 1. The chromosome sizes range from 2.55 Mbp to 2.67 Mbp, with G+C contents of 40.8 to 40.9%. We found seven to 10 plasmids (per strain) with G+C contents from 34.7 to 43.9%. Plasmid sizes range from 7,030 bp to 70,980 bp, resulting in overall genome sizes of 2.78 to 2.85 Mbp. The analysis of RAST-annotated genomes resulted in an *L. backii* core genome with 1,924 gene families and a pangenome with 2,889 gene families. The chromosomes encode five complete rRNA operons and 66 to 68 tRNAs.

The analysis of all five *L. backii* genomes revealed the presence of the same brewery-specific (99% sequence similarity, 99% coverage to each other) and plasmid-encoded fatty acid biosynthesis cluster as found in case of *Pediococcus damnosus* (6). Similarly, *L. backii* encodes an incomplete chromosomal fatty acid biosynthesis. Long-chain fatty acids are scarce in beer (13), while it was shown that the ability to produce fatty acids *de novo* is essential for *P. damnosus* growth in beer (6). The availability of these five *L. backii* genome sequences provides the basis for consecutive

TABLE 1 Strain characteristics, sequencing statistics, genome information, and accession numbers<sup>a</sup>

Strain	Source	BioSample no.	Accession no.	Avg coverage of HGAP assembly (×)	Size (Mbp)	No. of contigs	G+C content (%)	No. of PEGs	No. of CDSs
TMW 1.1988	Light wheat beer	SAMN04505726	CP014623 to CP014633	121	2.82	11	40.8	2,671	2,495
TMW 1.1989	Beer	SAMN04505727	CP014873 to CP014880	89	2.85	8	40.8	2,646	2,496
TMW 1.1991	Brewery environment	SAMN04505728	CP014881 to CP014889	99	2.82	9	40.7	2,590	2,437
TMW 1.1992	Brewery environment <sup>b</sup>	SAMN04505729	CP014890 to CP014898	109	2.78	9	40.8	2,621	2,450
TMW 1.2002	Brewery environment <sup>b</sup>	SAMN04505730	CP014899 to CP014906	168	2.84	8	40.7	2,653	2,478

<sup>a</sup> All strains (BioSamples) have beer spoilage ability and have been isolated from German breweries. All BioSamples are part of the BioProject PRJNA290141. Accession numbers are listed for all contigs of each whole genome (as range). Number of contigs are from chromosome plus plasmids and partial plasmids (only the case for TMW 1.1992). PEG, protein-encoding genes based on RAST annotation; CDS, coding sequences (coding) based on NCBI PGAP.

<sup>b</sup> TMW 1.1992 and TMW 1.2002 are from the same brewery.

analyses (e.g., transcriptomics and plasmid curing experiments), with the objective to derive novel lifestyle genes of beer-spoiling *L. backii*. It will further help understand the role of plasmids for LAB niche adaptation.

**Accession number(s).** The five complete *L. backii* genomes have been deposited in DDBJ/EMBL/GenBank under the accession numbers stated in Table 1.

## FUNDING INFORMATION

This work, including the efforts of Rudi F. Vogel, was funded by Allianz Industrie Forschung (AiF) (Aif 17576N).

Part of this work was funded by the German Ministry of Economics and Technology (via AiF) and the Wifoe (Wissenschaftsförderung der Deutschen Brauwirtschaft e.V., Berlin) in project AiF 17576N. None of the funding sources had any influence on the study design, the collection, analysis, and interpretation of data, the writing of the report, or the decision to submit the article for publication.

## REFERENCES

- Vriesekoop F, Krahl M, Hucker B, Menz G. 2012. 125th anniversary review: bacteria in brewing: the good, the bad and the ugly. *J Inst Brew* 118:335–345. <http://dx.doi.org/10.1002/jib.49>.
- Suzuki K. 2011. 125th anniversary review: microbiological instability of beer caused by spoilage Bacteria. *J Inst Brew* 117:131–155. <http://dx.doi.org/10.1002/j.2050-0416.2011.tb00454.x>.
- Geissler AJ, Behr J, von Kamp K, Vogel RF. 2016. Metabolic strategies of beer spoilage lactic acid bacteria in beer. *Int J Food Microbiol* 216:60–68. <http://dx.doi.org/10.1016/j.ijfoodmicro.2015.08.016>.
- Bohak I, Thelen K, Beimfohr C. 2006. Description of *Lactobacillus backii* sp. nov., an obligate beer-spoiling bacterium. *Monatsschrift für Brauwissenschaft* 59:78–82.
- Suzuki K. 2015. Gram-positive spoilage bacteria in brewing, p 141–174. *In* Hill AE (ed), *Brewing microbiology managing microbes, ensuring quality and valorising waste*. Woodhead publishing series in food science, technology, and nutrition, Woodhead Publishing, Cambridge, United Kingdom.
- Behr J, Geissler AJ, Schmid J, Zehe A, Vogel RF. 2016. The identification of novel diagnostic marker genes for the detection of beer spoiling *Pediococcus damnosus* strains using the BLAST diagnostic gene finder. *PLoS One* 11:e0152747. <http://dx.doi.org/10.1371/journal.pone.0152747>.
- Eid J, Fehr A, Gray J, Luong K, Lyle J, Otto G, Peluso P, Rank D, Baybayan P, Bettman B, Bibillo A, Bjornson K, Chaudhuri B, Christians F, Cicero R, Clark S, Dalal R, Dewinter A, Dixon J, Foquet M, Gaertner A, Hardenbol P, Heiner C, Hester K, Holden D, Kearns G, Kong X, Kuse R, Lacroix Y, Lin S, Lundquist P, Ma C, Marks P, Maxham M, Murphy D, Park I, Pham T, Phillips M, Roy J, Sebra R, Shen G, Sorenson J, Tomaney A, Travers K, Trulson M, Vieceli J, Wegener J, Wu D, Yang A, Zaccarin D. 2009. Real-time DNA sequencing from single polymerase molecules. *Science* 323:133–138. <http://dx.doi.org/10.1126/science.1162986>.
- Chin CS, Alexander DH, Marks P, Klammer AA, Drake J, Heiner C, Clum A, Copeland A, Huddleston J, Eichler EE, Turner SW, Korlach J. 2013. Nonhybrid, finished microbial genome assemblies from long-read SMRT sequencing data. *Nat Methods* 10:563–569.
- Angiuoli SV, Gussman A, Klimke W, Cochrane G, Field D, Garrity G, Kodira CD, Kyrpides N, Madupu R, Markowitz V, Tatusova T, Thomson N, White O. 2008. Toward an online repository of Standard Operating Procedures (SOPs) for (meta)genomic annotation. *Omic* 12: 137–141. <http://dx.doi.org/10.1089/omi.2008.0017>.
- Aziz RK, Bartels D, Best AA, DeJongh M, Disz T, Edwards RA, Formisano K, Gerdes S, Glass EM, Kubal M, Meyer F, Olsen GJ, Olson R, Osterman AL, Overbeek RA, McNeil LK, Paarmann D, Paczian T, Parrello B, Pusch GD, Reich C, Stevens R, Vassieva O, Vonstein V, Wilke A, Zagnitko O. 2008. The RAST server: Rapid Annotations using Subsystems Technology. *BMC Genomics* 9:75. <http://dx.doi.org/10.1186/1471-2164-9-75>.
- Overbeek R, Olson R, Pusch GD, Olsen GJ, Davis JJ, Disz T, Edwards RA, Gerdes S, Parrello B, Shukla M, Vonstein V, Wattam AR, Xia F, Stevens R. 2014. The SEED and the rapid annotation of microbial genomes using subsystems technology (RAST). *Nucleic Acids Res* 42: D206–D214. <http://dx.doi.org/10.1093/nar/gkt1226>.
- Vesth T, Lagesen K, Acar Ö, Ussery D. 2013. CMG-Biotools, a free workbench for basic comparative microbial genomics. *PLoS One* 8:e60120. <http://dx.doi.org/10.1371/journal.pone.0060120>.
- Preedy VR. 2009. *Beer in health and disease prevention*. Elsevier/ Academic Press, Amsterdam, The Netherlands.